

ZEBRAFISH

A MODEL ORGANISM IN BIOMEDICAL SCIENCES

MEDSCIEN9502

Team 6: Hamza Alsaied, Herthika Sivasuntharampillai, & Vanesa Berati



Agenda



1. INTRO TO
ZEBRAFISH

2. HISTORY OF
STUDY

3. ADVANTAGES &
LIMITATIONS

4. ZEBRAFISH
CASE STUDIES

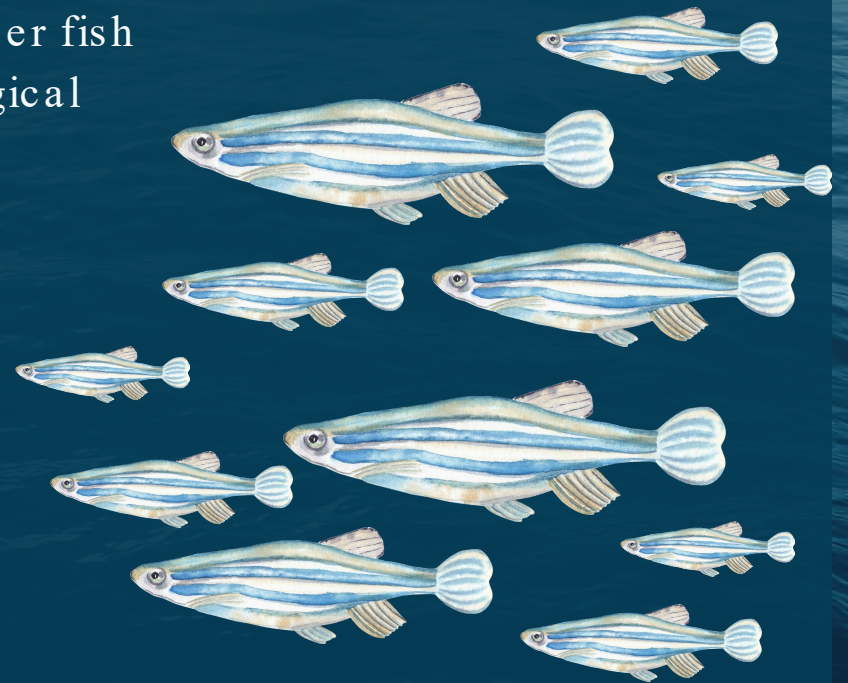


An aerial photograph of a coastline with a dark blue semi-transparent overlay. The overlay is a large rectangle that covers most of the image, leaving a thin border of the original photograph visible. The word "INTRODUCTION" is centered in white text within the dark blue area.

INTRODUCTION

Introduction

- Zebrafish (*Danio rerio*) are small freshwater fish
- Widely used as model organisms in biological and medical research
- Native to South Asia
- Key features:
 - Short generation time
 - High fertility
 - Rapid development
 - Eggs develop externally
 - Sexually mature by 12 weeks



An aerial photograph of a river delta, showing a complex network of water channels and land. The image is overlaid with a dark blue, semi-transparent rectangular box. The text "HISTORY AND KEY EVENTS" is centered within this box in a white, bold, sans-serif font.

HISTORY AND KEY EVENTS

History



First described by the British physician Francis Hamilton

1822

Early 20th Century

Popular as aquarium fish in Europe and the U.S

Charles W. Creaser published a paper describing the suitability of zebrafish for embryological research

1934

George Streisinger transitioned to working with zebrafish as a model for vertebrate genetics

1960s

1981

First major breakthrough in zebrafish research by developing a technique to generate homozygous diploid zebrafish through artificial parthenogenesis

Mutagenesis techniques were established

1983

History



The first study analyzing a neuronal necrosis mutant from mutagenesis experiments was published

1988

Charles Kimmel, a developmental biologist, published the first fate map for zebrafish

Results from these screens were published in Development journal

1996

Zebrafish Genome Project was initiated

1990s

Christiane Nüsslein-Volhard initiated preparations for large-scale mutagenesis screen

2001

Zebrafish have become a crucial model in human disease research

Large-scale mutagenesis screens were conducted

1993-1996

PRESENT

An aerial photograph of a coastline with a dark blue semi-transparent overlay. The text is centered within the overlay.

KEY ADVANTAGES & LIMITATIONS

Advantages

Space & cost efficiency

High genetic homology to humans

Less invasive and more reliable drug administration

Conserved physiology

Versatile application of developmental stages

Genetic tools

Abundant reproduction & fast development

Small brains

(Chia et al., 2020)
(Stewart et al., 2014)
(Vaz et al., 2018)

Limitations

Water-soluble pharmaceuticals strongly recommended

Physiological variability between species

Poorly understood pharmacodynamic and pharmacokinetic properties

Larvae do not have a BBB

Not as many well characterized strains

Complex behaviors in development

Inbreeding issues

Differences in brain regions

(Chia et al., 2020)
(Stewart et al., 2014)
(Vaz et al., 2018)

A microscopic image of zebrafish tissue, showing a complex network of cells and structures. The image is overlaid with a dark blue semi-transparent rectangle that contains the title text.

ZEBRAFISH IN BIOMEDICAL RESEARCH

Genetics & Development

- zebrafish reference genome: GRCz11
- *in vivo* model for studying causal genes and mechanisms underlying human disease pathogenesis
- amenable to genetic manipulations
 - CRISPR/Cas9
 - next-generation sequencing (NGS) & long-read nanopore sequencing
 - functional validation of GWAS findings
- optogenetics and fluorescent markers for tracking cell processes over time
 - non-invasive, real-time imaging of internal structures



The Company of Biologists (2015)

Kar & Subbiah (2013)
Howe et al. (2013)
Choi et al. (2021)
Chernyavskaya et al. (2022)

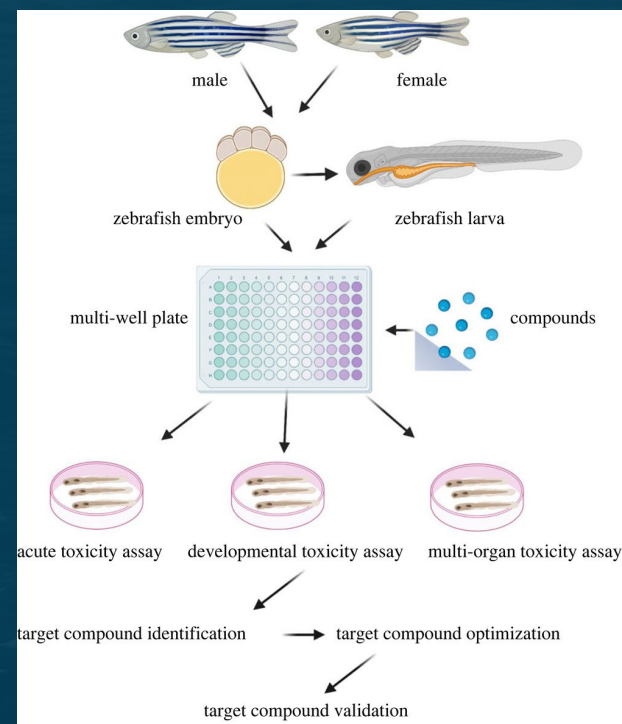
Drug Development & Toxicology

High-throughput Drug Screening

- Zebrafish embryos are permeable to small molecules, enabling rapid compound testing.
- *in vivo* approaches for phenotype-based screening
 - study signalling cascades using chemical inhibitors
 - assess toxicity and target identification
 - apply AI-driven imaging to track and visualize tagged compounds

Precision Medicine applications:

- new molecular targets for diagnostic and therapeutic strategies



Burke (2016)

Choi et al. (2021)

Dash & Patnaik (2023)

Angom & Nakka (2024)

Adapted from: Dash & Patnaik (2023)

NEUROSCIENCE

NEURODEVELOPMENTAL RESEARCH

Studying Neuronal Development

- Well-conserved nervous system structures with humans
- Model for studying neurogenesis, neuron mapping, and neuroendocrine functions.
- Transparent larval stage allows *in vivo* monitoring of motor neuron pathways and degeneration.

Chia et al. (2022)
Dash & Patnaik (2023)
Angom & Nakka (2024)

Howard Hughes Medical Institute (2013)

NEUROSCIENCE

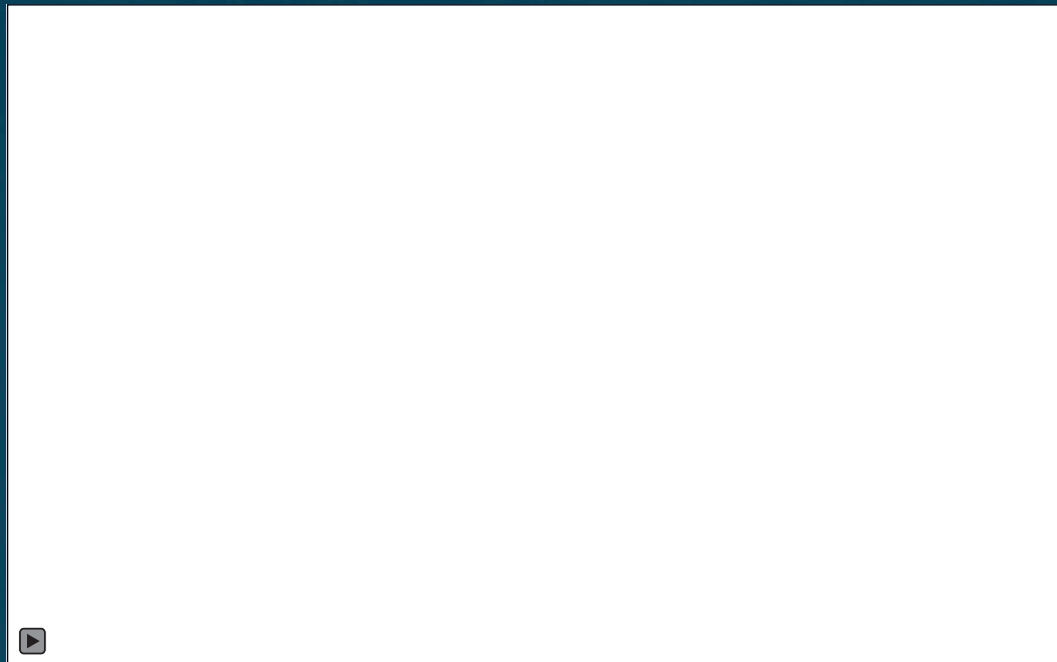
MODELLING NEURODEGENERATIVE DISEASES

Application in ALS, Parkinson's, Alzheimer's, Huntington's

- Models recreate key pathology features (e.g., protein aggregation, neuron degeneration)

Case Study: ALS and FUS Mutation Studies

- Used to uncover pathological mechanisms like neuron degeneration in ALS (Lebedeva et al., 2016).



Howard Hughes Medical Institute (2013)

Choi et al. (2021)

Chia et al. (2022)

COMPLEX DISEASES

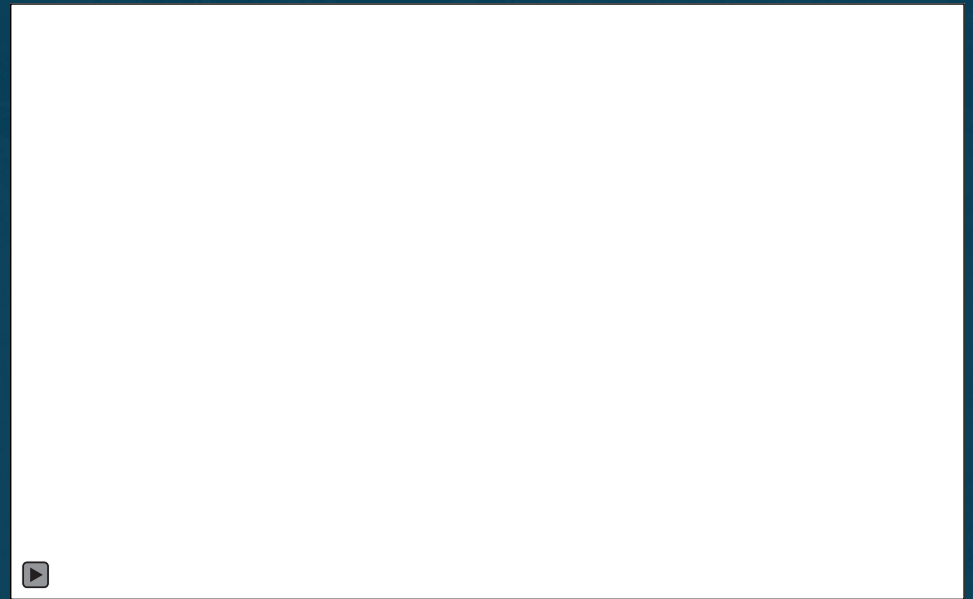
CARDIOVASCULAR RESEARCH

Heart Disease and Regeneration Models

- Zebrafish hearts mimic human cardiac processes, aiding studies in CVD.
- High regenerative capacity of zebrafish cardiomyocytes provides insights into cardiac regeneration.

Doxorubicin Cardiotoxicity Model

- Identification of cardioprotective agents
 - visnagin (VIS) and diphenylurea (DPU)
- found to combat chemotherapy-induced heart damage (Lui et al., 2014)



Nightsea (2014)

Kar & Subbiah (2013)
Dash & Patnaik (2023)
Angom & Nakka (2024)

COMPLEX DISEASES

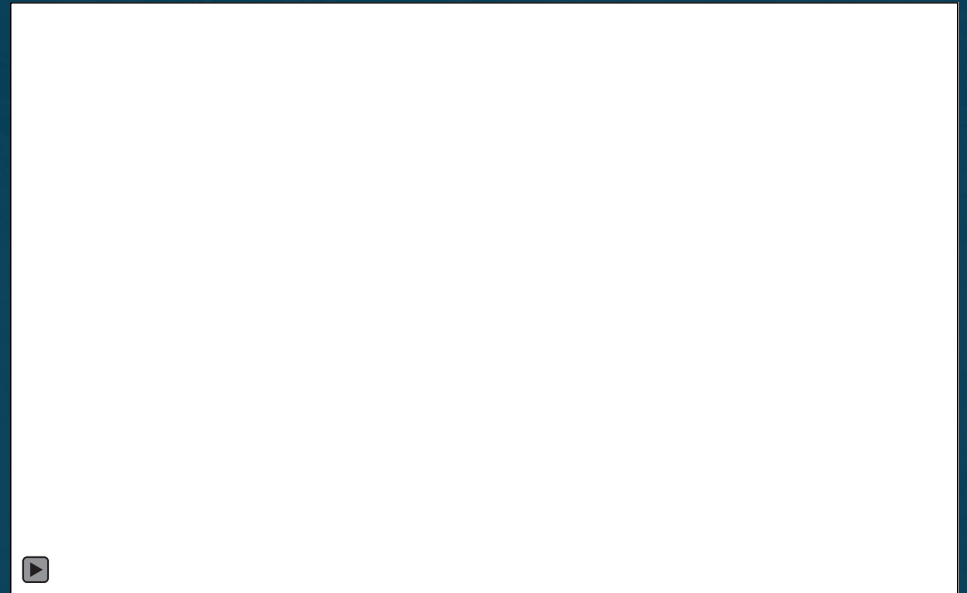
ONCOLOGY RESEARCH

Cancer Modeling and Drug Screening

- Tumour growth visualization, drug screening, and toxicity testing.

Success in Kinase Inhibitor Identification

- Found compounds targeting cell proliferation and migration in melanoma and leukemia (Hason et al., 2023).
- high sensitivity and accuracy for quantifying cancer cell growth
 - valuable model for pre-clinical drug screening.



Cancer Research UK (2014)

Dash & Patnaik (2023)

Conclusions

Zebrafish

A versatile and powerful model organism in biomedical research due to their genetic and physiological similarity to humans.

Advantage

Zebrafish's transparency, high fecundity, and genetic manipulability make them especially useful for developmental biology, genetics, disease modelling, and drug discovery.

Future

CRISPR, NGS, and AI-driven imaging enhance their role in personalized medicine and drug discovery, furthering insights into human diseases, therapeutic targets, and regenerative medicine applications.

THANK YOU
FOR LISTENING



References

Angom, R. S., & Nakka, N. M. R. (2024). Zebrafish as a Model for Cardiovascular and Metabolic Disease: The Future of Precision Medicine. *Biomedicines*, *12* (3), 693. <https://doi.org/10.3390/biomedicines12030693>

Ankeny, R. A., Chadrevian, S. de, Gritsman, K., Grunwald, D. J., Halpern, M. E., Kimmel, C. B., Müller-Wille, S., Mullins, M. C., Zhang, J., Bechtel, W., Benzer, S., Bisgrove, B. W., Bolker, J. A., Brand, M., Brenner, S., Burian, R. M., Canguilhem, G., Cheng, K. C., Creaser, C. W., ...Griesemer, J. R. (2012, February 9). Stages in the development of a model organism as a platform for mechanistic models in developmental biology: Zebrafish, 1970–2000. *Studies in History and Philosophy of Science Part C: Studies in History and Philosophy of Biological and Biomedical Sciences*. <https://www.sciencedirect.com/science/article/abs/pii/S1369848611001403>

Burke, E. (2016, August 9). Why Use Zebrafish to Study Human Diseases? *The Intramural Research Program*. <https://irp.nih.gov/blog/post/2016/08/why-use-zebrafish-to-study-human-diseases>

Cancer Research UK. (2014, July 24). Fluorescent Fish and Skin Cancer | Cancer Research UK [Video]. YouTube. <https://www.youtube.com/watch?v=LaInISRESrU>

Chernyavskaya, Y., Zhang, X., Liu, J., & Blackburn, J. (2022). Long-read sequencing of the zebrafish genome reorganizes genomic architecture. *BMC Genomics* *23* , 116. <https://doi.org/10.1186/s12864-022-08349-3>

Chia, K., Klingseisen, A., Sieger, D., & Priller, J. (2022). Zebrafish as a model organism for neurodegenerative disease. *Frontiers in Molecular Neuroscience*, *15*, 940484. <https://doi.org/10.3389/fnmol.2022.940484>

Chia, S. J., Tan, E.-K., & Chao, Y.-X. (2020). Historical Perspective: Models of Parkinson's Disease. *International Journal of Molecular Sciences*, *21* (7), 2464. <https://doi.org/10.3390/ijms21072464>

Choi, T.-Y., Choi, T.-I., Lee, U.-R., Choe, S.-K., & Kim, C.-H. (2021). Zebrafish as an animal model for biomedical research. *Experimental & Molecular Medicine*, *53* , 310-317. <https://doi.org/10.1038/s12276-021-00571-5>

The Company of Biologists. (2015, February 24). Beautiful imaging of zebrafish development [Video]. YouTube. <https://www.youtube.com/watch?v=yk7TWOtrpHM>

Dash, S. N., & Patnaik, L. (2023). Flight for fish in drug discovery: a review of zebrafish-based screening of molecules. *Biology Letters*, *19* , 20220541. <https://doi.org/10.1098/rsbl.2022.0541>

References

Hason, M., Jovicic, J., Vonkova, I., Bojic, M., Simon -Vermot, T., White, R. M., & Bartunek, P. (2022). Bioluminescent Zebrafish Transplantation Model for Drug Discovery. *Frontiers in Pharmacology*, *13*, 893655. <https://doi.org/10.3389/fphar.2022.893655>

Howard Hughes Medical Institute. (2013, March 18). Whole-brain imaging of neuronal activity in a larval zebrafish [Video]. YouTube. <https://www.youtube.com/watch?v=lpAwkek6DI>

Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Collins, J. E., Humphray, S., McLaren, K., Matthews, L., McLaren, S., Sealy, I., Caccamo, M., Churcher, C., Scott, C., Barrett, J. C., Koch, R., Rauch, G. J., White, S., ...Stemple, D. L. (2013). The zebrafish reference genome sequence and its relationship to the human genome. *Nature*, *496* (7446), 498–503. <https://doi.org/10.1038/nature12111>

JP;, B. (2002, January 1). The zebrafish: A new model organism for Integrative Physiology. American journal of physiology. Regulatory, integrative and comparative physiology. <https://pubmed.ncbi.nlm.nih.gov/11742817/>

Kar, B., & Subbiah, S. (2013). Zebrafish: An in Vivo Model for the Study of Human Diseases. *International Journal of Genetics and Genomics*, *1* (1), 6-11. <https://doi.org/10.11648/j.ijgg.20130101.12>

Lebedeva, S., de Jesus Domingues, A. M., Butter, F., & Ketting, R. F. (2017). Characterization of genetic loss-of-function of Fus in zebrafish. *RNA Biology*, *14* (1), 29–35. <https://doi.org/10.1080/15476286.2016.1256532>

Liu, Y., Asnani, A., Zou, L., Bentley, V. L., Yu, M., Wang, Y., Dellaire, G., Sarkar, K. S., Dai, M., Chen, H. H., Sosnovik, D. E., Shin, J. T., Haber, D. A., Berman, J. N., Chao, W., & Peterson, R. T. (2014). Visnagin protects against doxorubicin-induced cardiomyopathy through modulation of mitochondrial malate dehydrogenase. *Science Translational Medicine*, *6* (266). https://doi.org/10.1126/SCITRANSLMED.3010189/SUPPL_FILE/6-266RA170_SM.PDF

Nightsea. (2014, September 23). Green-fluorescent transgenic zebrafish heart beating [Video]. YouTube. <https://www.youtube.com/watch?v=fYMEOEN7ANM&t=52s>

Stewart, A. M., Braubach, O., Spitsbergen, J., Gerlai, R., & Kalueff, A. V. (2014). Zebrafish models for translational neuroscience research: From tank to bedside. *Trends in Neurosciences*, *37*(5), 264–278. <https://doi.org/10.1016/j.tins.2014.02.011>